



Dietary Phenolics as Reactive Carbonyl Scavengers: Potential Impact on Human Health and Mechanism of Action

Chi-Tang Ho¹, Mingfu Wang²

¹Department of Food Science, Rutgers University, New Brunswick, NJ, USA.

²School of Biological Sciences, The University of Hong Kong, Hong Kong, P. R. China.

ABSTRACT

Previous studies have demonstrated that accumulation of reactive carbonyl compounds in human tissue will accelerate the vascular damage in both diabetes and uremia. Moreover, advanced glycation progressively and irreversibly modify the proteins over time and yield advanced glycation end products (AGEs). AGEs are thought to contribute to the development of diabetes mellitus and its complications. Therefore, we propose a novel approach to decrease the levels of dicarbonyl compounds by direct trapping of dietary polyphenolic compounds, and consequently, inhibit the formation of AGEs and prevent the development of diabetic complications and age-related diseases.

Key words: Advanced glycation end products, Diabetes, Dietary phenolic compounds, Epigallocatechin-3-gallate, Reactive carbonyl species

REACTIVE CARBONYL SPECIES, ADVANCED GLYCATION END PRODUCTS, AND AGE-RELATED DISEASES

Non-enzymatic modifications of proteins have been implicated in the pathogenesis of diabetes, atherosclerosis, neurodegenerative diseases, and normal aging.^[1,2] The modifications can arise from direct exposure to reactive oxygen, chlorine, or nitrogen species, and from reaction with low-molecular-weight reactive carbonyl species (RCS), which originate from a multitude of mechanistically related pathways, like glycation, sugar autoxidation, lipid peroxidation, and UV-photodamage. The

accumulation of various RCS such as glyoxal (GO), methylglyoxal (MGO) derived from carbohydrates or lipids, as well as their subsequently induced protein modifications are proposed to constitute a state of “carbonyl stress.”^[3] These RCS are responsible for the formation of advanced glycation end products (AGEs) and advanced lipoxidation end products (ALEs), and their roles in the development of various aging-related diseases have been increasingly recognized.^[4] Higher levels of RCS were observed in the plasma of diabetes patients than in the plasma of healthy people.^[5] Therefore, decreasing the levels of dicarbonyl compounds, and consequently, inhibiting the formation of AGEs would be a useful approach to prevent the development of diabetic complications. There is thus a

Correspondence to:

Prof. Chi-Tang Ho, Department of Food Science, Rutgers University, 65 Dudley Road, New Brunswick, NJ 08901, USA. Tel: 848-932-5553.

E-mail: ho@aesop.rutgers.edu

DOI: 10.4103/2225-4110.114892

prompt need to develop effective strategies to protect from RCS-associated pathogenic conditions, and this will remain one of the major research directions that merit global intention. A very limited number of chemical agents have been found to suppress or prevent excessive generation and accumulation of cellular RCS, and their therapeutic potential has been recognized only recently. These compounds exert their action by interfering with different phases of the reaction cascades, such as by acting as antioxidants, by chelating metal ions, or by directly trapping RCS. As free radical-mediated and oxidative reactions are known to be involved in the process of glycation and lipoxidation, it is not a surprise that antioxidants may be effective inhibitors of glycation and/or lipoxidation in *in vitro* assays. However, numerous clinical trials have failed to provide conclusive evidence for the efficacy of antioxidant therapy in several chronic diseases. These findings have created doubt about the effectiveness of chemical agents that behave solely as antioxidants in alleviating carbonyl stress. An integration of these previous findings and information regarding the formation pathways of RCS, AGEs, and ALEs has enabled us to put forward the hypothesis that chemical agents possessing dual mechanisms of action, namely antioxidant and RCS-trapping activities, are likely to be more promising candidates for developing into disease-preventive agents/pharmaceutical leads for age-related diseases.

TRAPPING OF RCS BY DIETARY PHYTOCHEMICALS

In our studies as well as those of other laboratories, various natural extracts and phytochemicals have been evaluated for their effects on RCS-induced modification of protein structure. Yet, only a very limited number of natural products have demonstrated RCS-trapping capacity.^[6-15] In a previous study, we found that epigallocatechin-3-gallate (EGCG) could rapidly trap both MGO and GO under neutral or alkaline conditions. Our data showed that EGCG was more reactive than lysine and arginine in terms of trapping MGO or GO, indicating that EGCG has the potential to compete with lysine and arginine *in vivo* and, therefore, prevent the formation of AGEs. In addition, we also found that EGCG was more reactive at trapping MGO than the pharmaceutical agent, aminoguanidine, which has been shown to inhibit the formation of AGEs by trapping of reactive dicarbonyl compounds *in vivo*.^[16]

We have purified three major products from the reaction between EGCG and MGO at a 3:1 mole ratio. Their structures were identified as two mono-MGO adducts and one di-MGO adduct of EGCG with the MGO conjugated at positions 6 and 8 of the EGCG A-ring [Figure 1]. Our results clearly indicate that the major active site of EGCG is at positions 6 and 8 of the A-ring and that the gallate ring does not play an important role in the trapping of reactive dicar-

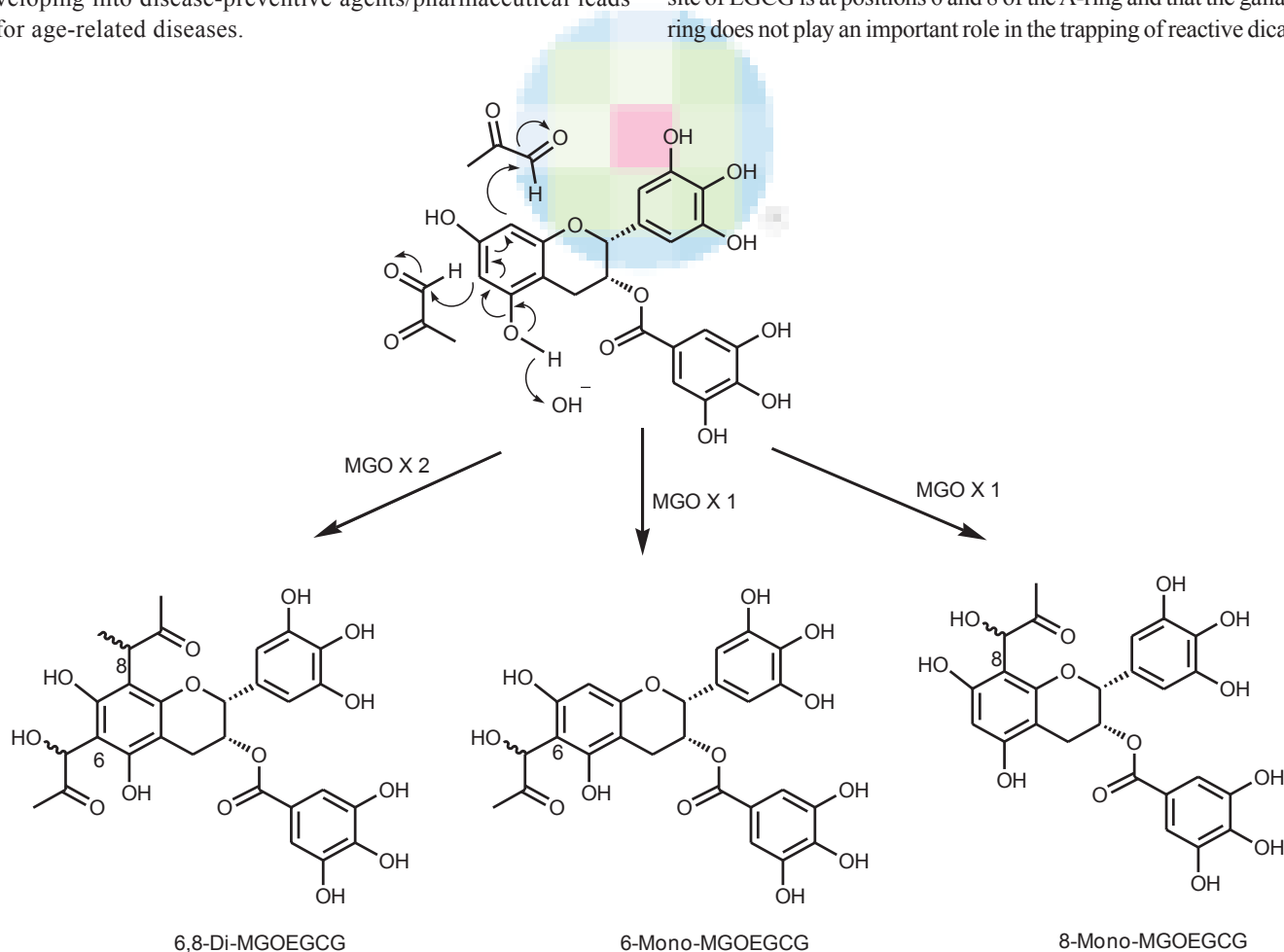


Figure 1. Adducts of methylglyoxal and epigallocatechin-3-gallate

bonyl species. The slightly alkaline pH can increase the nucleophilicity at positions 6 and 8 of the A-ring of EGCG, facilitating the addition of MGO at these two positions to form mono- and di-MGO adducts.

Besides EGCG, catechin, epicatechin, theaflavin,^[6] proanthocyanidins,^[9] phloretin, phloridzin,^[10] genistein,^[12] curcumin,^[14] and a stilbene glucoside from *Polygonum multiflorum* Thunb.^[11] can effectively trap MGO. Therefore, these compounds represent a new group of 1,2-dicarbonyl scavenging agents. However, these hypotheses must be proven by *in vitro* and *in vivo* studies with the AGEs being accurately analyzed. In addition, different from traditional views on drugs (most drugs elicit their effects via transient interactions with membrane-spanning receptors that modulate cellular signaling pathways), ideally, the carbonyl scavengers should show minimal activity toward drug receptors, thus minimizing unwanted pharmacological effects. Rather, the administration of carbonyl scavengers should proceed in the expectation that they rapidly sequester carbonyl species in cells, thus blocking the adduction of macromolecules and any downstream damages. Whether these phenolic compounds can selectively perform this function also demands further study.

REFERENCES

1. Singh R, Barden A, Mori T, Beilin L. Advanced glycation end-products: A review. *Diabetologia* 2001;44:29-46.
2. Baynes JW. The role of AGEs in aging: Causation or correlation. *Exp Gerontol* 2001;36:1527-37.
3. Baynes JW, Thorpe SR. Role of oxidative stress in diabetic complications: A new perspective on an old paradigm. *Diabetes* 1999;48:1-9.
4. Baynes JW, Thorpe SR. Glycoxidation and lipoxidation in atherogenesis. *Free Rad Biol Med* 2000;28:1708-16.
5. Khuwawar MY, Kandhro AJ, Khand FD. Liquid chromatographic determination of glyoxal and methylglyoxal from serum of diabetic patients using meso-stilbenediamine as derivatizing agent. *Anal Lett* 2006;39:2205-15.
6. Lo CY, Li S, Tan D, Pan MH, Sang S, Ho CT. Trapping reactions of reactive carbonyl species with tea polyphenols in simulated physiological conditions. *Mol Nutr Food Res* 2006;50:1118-28.
7. Sang S, Shao X, Bai N, Lo CY, Yang CS, Ho CT. Tea polyphenol (-)-epigallocatechin-3-gallate: A new trapping agent of reactive dicarbonyl species. *Chem Res Toxicol* 2007;20:1862-70.
8. Tan Di, Wang Y, Lo CY, Ho CT. Methylglyoxal: Its presence and potential scavengers. *Asia Pac J Clin Nutr* 2008;17:257-60.
9. Peng X, Cheng KW, Ma J, Chen B, Ho CT, Chen F, *et al.* Cinnamon bark proanthocyanidins as reactive carbonyl scavengers to prevent the formation of advanced glycation endproducts. *J Agric Food Chem* 2008;56:1907-11.
10. Shao X, Bai N, He K, Ho CT, Yang CS, Sang S. Apple polyphenols, phloretin and phloridzin: New trapping agents of reactive dicarbonyl species. *Chem Res Toxicol* 2008;21:2042-50.
11. Lv L, Shao X, Wang L, Huang D, Ho CT, Sang S. Stilbene glucoside from *Polygonum multiflorum* Thunb.: A novel natural inhibitor of advanced glycation end product formation by trapping of methylglyoxal. *J Agric Food Chem* 2010;58:2239-45.
12. Lv L, Shao X, Chen H, Ho CT, Sang S. Genistein inhibits advanced glycation end products formation by trapping of methylglyoxal. *Chem Res Toxicol* 2011;24:579-86.
13. Wang Y, Ho CT. Flavour chemistry of methylglyoxal and glyoxal. *Chem Soc Rev* 2012;41:4140-9.
14. Hu TY, Liu CL, Chyau CC, Hu ML. Trapping of methylglyoxal by curcumin in cell-free systems and in human umbilical vein endothelial cells. *J Agric Food Chem* 2012;60:8190-6.
15. Lo CY, Hsiao WT, Chen XY. Efficiency of trapping methylglyoxal by phenols and phenolic acid. *J Food Sci* 2011;76:H90-6.
16. Nilsson BO. Biological effects of aminoguanidine: An update. *Inflammation Res* 1999;48:509-15.